MALT lymphoma in labial salivary gland biopsy from Sjögren syndrome: importance of follow-up in early detection

A. Keszler, DDS, PhD, L. I. Adler, DDS, PhD, M. S. Gandolfo, DDS, PhD, P. A. Masquijo Bisio, DDS, A. C. Smith, DDS, C. F. Vollenweider, MD, A. M. Heidenreich, MD, G. de Stefano, MD, M. V. Kambo, BA, D. P. Cox, DDS, M. Narbaitz, MD, and H. E. Lanfranchi, DDS, PhD

Buenos Aires, Argentina

Mucosa-associated lymphoid tissue (MALT) lymphomas are known to occur in Sjögren syndrome (SS) patients, but reported cases in labial salivary glands (LSG) are rare. We report a case of 60-year-old female patient with SS who developed MALT lymphoma in the labial salivary glands during a 2-year time interval when she was participating in the Sjögren’s International Clinical Collaborative Alliance, an ongoing longitudinal multisite observational study funded by the National Institutes of Health of the United States. At follow-up exam, LSG biopsy showed atypical diffuse infiltration by mononuclear cells of variable size and atypical nuclei affecting the whole specimen with destruction of glandular architecture, leading to a diagnosis of B-cell MALT lymphoma. Computerized tomography and bone marrow biopsy failed to show additional evidence of disease. Clinical, serologic, ocular, histologic and immunohistochemical findings are presented. A “watch and wait” policy was adopted with regular examinations. (Oral Surg Oral Med Oral Pathol Oral Radiol 2013;115:e28-e33)

Primary Sjögren syndrome (SS) is a chronic autoimmune disease affecting the exocrine glands and other organs and producing circulating antibodies. Lymphocytic infiltration of the salivary and lacrimal glands leads to the progressive loss of the glandular parenchyma with decrease of function resulting in chronic salivary hypofunction and keratoconjunctivitis sicca, two of the major components of the syndrome. The oral dryness increases the risk of caries, and the mucosa is more prone to candidiasis and/or bacterial infections. Hernández et al. reported a 37% prevalence of atrophic chronic candidiasis in 246 patients with SS. Frequently the gastrointestinal tract is compromised, with the immunophenotype of the lymphoid cells that infiltrate this mucosa being very similar to that found in the minor salivary glands. Also gastric infection by Helicobacter pylori (Hp) has been reported in patients with SS. Aragona et al. communicated a significantly higher prevalence of antibodies against Hp and its heat-shock protein 60 (HSP60) in the serum of patients with SS than that found in a group of patients with other autoimmune diseases and healthy control subjects. Those authors concluded that the hypothetic role of HSP60 in the development of the immune response in both primary and secondary SS seems to be linked to the prevalence rate of Hp infection. The most likely mode of transmission of this bacterium is from person to person, mainly orally, because Hp DNA is present in both saliva and dental plaque.

The evolution of SS is usually indolent, although affected patients have a higher risk of developing non-Hodgkin lymphoma (NHL) than the general population, estimated to be 44 times greater. This is the most serious complications of SS. The incidence of lymphomas described by Voulgarelis et al. in patients with SS is 4.3%, and according to Sutcliffe et al. it varies between 5% and 10%. In most cases, they are extranodal B-cell lymphomas of low grade and mucosa-associated lymphoid tissue (MALT) lymphoma, also known as extranodal marginal zone B-cell lymphoma (MZL). This kind of lymphoma has unique clinicopathologic characteristics.
The World Health Organization divides MZLs into 3 subtypes: a) extranodal MZL (MALT-type lymphoma); b) splenic MZL; and c) nodal MZL (with or without monocytoid cells).\textsuperscript{10} The majority of the extranodal lymphomas affect mainly the digestive system and are originated from the lymphoid tissue of the mucosa and acquired secondary to a chronic inflammatory process. This inflammatory process produces the recruitment of type T and B lymphocytes to the affected site, and acquires the structure of organized lymphoid tissue, similar to normal MALT found in Peyer patches.\textsuperscript{9,10} In the gastric mucosa, MALT lymphoma is frequently associated with Hp infection.\textsuperscript{11} In the same way, cutaneous MZL has been associated with infection by \textit{Borrelia burgdorferi}, ocular adnexa MZL with \textit{Chlamydia psittaci}, small bowel MZL with \textit{Campylobacter jejuni}, and MZL of the salivary glands with hepatitis C virus.\textsuperscript{12} In addition, autoimmune diseases, such as SS, are a predisposing factor for its development in specific sites of the head and neck, including salivary glands.\textsuperscript{13,15}

In SS, MALT lymphoma originates mainly in the major salivary glands, particularly the parotid.\textsuperscript{12-25} Reported cases in the minor salivary glands are very rare. Speight et al.\textsuperscript{26} in their study of lower lip minor labial salivary glands reported the early detection of 4 cases of MALT lymphoma by in situ hybridization for kappa and lambda light chain mRNA in 14 cases of SS; van Mello et al.\textsuperscript{27} reported 3 cases in the same location, and Sakuma et al.\textsuperscript{28} reported 1 case in minor salivary glands of the hard palate. Table 1 summarizes MALT lymphoma cases of salivary glands of patients with SS found in the English-language literature in the past 15 years.

### Table 1. MALT lymphomas in salivary glands of patients with Sjögren syndrome

<table>
<thead>
<tr>
<th>Authors</th>
<th>Year</th>
<th>n</th>
<th>Localization</th>
<th>Diagnostic criteria*</th>
</tr>
</thead>
<tbody>
<tr>
<td>Speight et al.\textsuperscript{26}</td>
<td>1994</td>
<td>4</td>
<td>Minor labial salivary glands</td>
<td>κ and λ light chain; in situ hybridization for mRNA</td>
</tr>
<tr>
<td>Royer et al.\textsuperscript{14}</td>
<td>1997</td>
<td>3</td>
<td>Parotid</td>
<td>REAL classification; CD20, CD3, κ and λ light chain, Bcl2</td>
</tr>
<tr>
<td>Nishimura et al.\textsuperscript{15}</td>
<td>2000</td>
<td>1</td>
<td>Parotid</td>
<td>Immunohistochemical; Southern blotting immunoglobulin gene rearrangement</td>
</tr>
<tr>
<td>Biasi et al.\textsuperscript{16}</td>
<td>2001</td>
<td>6</td>
<td>Parotid and submaxillary</td>
<td>REAL classification; histomorphology and immunohistochemical study‡</td>
</tr>
<tr>
<td>Queneau et al.\textsuperscript{17}</td>
<td>2002</td>
<td>1</td>
<td>Parotid</td>
<td>CD20; Southern blot</td>
</tr>
<tr>
<td>Klussmann et al.\textsuperscript{18}</td>
<td>2003</td>
<td>1</td>
<td>Parotid</td>
<td>‡</td>
</tr>
<tr>
<td>Dunn et al.\textsuperscript{20}</td>
<td>2004</td>
<td>4</td>
<td>Parotid</td>
<td>WHO classification; CD3, CD20, CD-45RO, κ and λ light chain</td>
</tr>
<tr>
<td>Ambrosetti et al.\textsuperscript{12}</td>
<td>2004</td>
<td>15</td>
<td>Parotid</td>
<td>REAL/WHO classification; CD20, CD5, CD21, CD10, CD3, κ and λ light chain</td>
</tr>
<tr>
<td>Streubel et al.\textsuperscript{19}</td>
<td>2004</td>
<td>15</td>
<td>Parotid and Submaxillary</td>
<td>WHO classification; reverse transcription PCR; fluorescence in situ hybridization</td>
</tr>
<tr>
<td>van Mello et al.\textsuperscript{27}</td>
<td>2005</td>
<td>3</td>
<td>Minor labial salivary GLANDS</td>
<td>Monoclonal κ and λ light chain; CD 43, CD 20, CD3</td>
</tr>
<tr>
<td>Sakuma et al.\textsuperscript{28}</td>
<td>2006</td>
<td>1</td>
<td>Minor salivary gland hard palate</td>
<td>CD20, CD3, CD5, CD10, CD23, cyclin D1, immunoglobulin light chain</td>
</tr>
<tr>
<td>Arcaini et al.\textsuperscript{21}</td>
<td>2006</td>
<td>4†</td>
<td>Parotid</td>
<td>WHO classification; immunophenotypic profile was used to exclude other type of indolent low-grade B-cell lymphomas‡</td>
</tr>
<tr>
<td>Lewis et al.\textsuperscript{22}</td>
<td>2007</td>
<td>2</td>
<td>Parotid and submaxillary</td>
<td>CD20, CD5, CD10, CD23, cyclin D1, κ light chain</td>
</tr>
<tr>
<td>Roh et al.\textsuperscript{24}</td>
<td>2008</td>
<td>1†</td>
<td>Parotid</td>
<td>REAL classification; CD3, CD5, CD10, CD20, CD23, CD43, CD-45RO, CD79a, κ and λ light chain, Bcl2, Bcl6, Ki-67</td>
</tr>
<tr>
<td>Suh et al.\textsuperscript{13}</td>
<td>2008</td>
<td>2</td>
<td>Parotid and submaxillary (both)</td>
<td>WHO classification; CD3,CD5,CD10,CD20,CD21,CD43,CD79a,cyclinD1,κandλlightchain,Bcl2,Bcl6,Ki-67</td>
</tr>
<tr>
<td>Hu et al.\textsuperscript{23}</td>
<td>2009</td>
<td>6</td>
<td>Parotid</td>
<td>WHO classification; CD3, CD20, CD10, Bcl6, κ and λ light chain, IgM, IgG, IgA</td>
</tr>
</tbody>
</table>

*Classification and immunohistochemical markers used.
†Only patients with SS were considered.
‡Nonspecified.
lip; and blood test determination of levels of rheumatoid factor (RF), IgG, IgA, IgM, C3, C4, antinuclear antibodies (ANA), anti-SSA and anti-SSB antibodies, and anti–hepatitis C antibodies. A follow-up assessment is carried out 2 years later with repeat of all studies.

CLINICAL CASE

A 60-year-old female patient presented at the Oral Medicine clinic of the School of Dentistry of the University of Buenos Aires in December 2004 with dry mouth symptoms and burning sensation. She reported xerophthalmia for 15 years and xerostomia for 4 years and had not used anticholinergic drugs. Clinical examination revealed marked dryness and atrophy of the buccal mucosa, with the presence of traumatic erosions associated with lack of moisture. Given these clinical characteristics, she was a candidate for and consented to be enrolled in the SICCA study. The initial oral examination was performed and revealed clinical manifestations of chronic candidiasis, confirmed by culture, and marked xerostomia. Saliva expressed from both parotid ducts was thick, and there was no expression from the submandibular ducts. The patient wore maxillary and mandibular complete dentures. Sialometry showed total unstimulated salivary flow rate of 0 mL/5 min (reference range [RR] 2 g/5 min) and a stimulated parotid salivary flow rate of 0 mL/5 min (RR 0.1 g/5 min).

At the rheumatologic examination, the patient stated not having pain and presented with mild asthenia. At the time of the visit the patient was taking 500 mg/d paracetamol and 0.25 mg 3 times a week alprazolam and using artificial tears. Ocular examination revealed: keratoconjunctivitis sicca demonstrated by Schirmer test: 0 mm; tear break-up time (BUT): 1 second; corneal staining with 0.5% fluorescein, with the only difference that filaments could be observed. The results were obtained for the Schirmer test, BUT, and conjunctival staining with 1% lissamine green for both eyes.

Blood test showed levels of RF, IgA, IgG, IgM, C3, and C4 within normal limits. The ANA was high at 1:1,280 (RR 1-40), with a speckled pattern, whereas the anti-SSA and anti-SSB antibodies were negative. Anti–hepatitis C was nonreactive. In addition, serum anti-Hp IgG antibodies, and secretory IgA antibodies were requested for evaluation of the presence of Hp. IgG was negative with a value of 2.57 (RR >15) and secretory IgA also was negative with a value of 18 (normal range 7-18).

Histopathology of the biopsied labial salivary glands showed multiple dense foci (n = 18) of typical lymphocytes with periductal and perivascular orientation and no evidence of germinal centers or lymphoepithelial lesions. The diagnosis was focal lymphocytic sialadenitis. The evaluated glandular area was 7.5 mm² and the calculated focus score was 8.5. Based on the working standard used at the time within the SICCA registry for patient management, the patient was informed that she had both the ocular and the oral components of SS.

The patient periodically returned to the Oral Medicine clinic for follow-up and management of her oral dryness and associated complications. In March 2007, the second evaluation was completed according to the project protocol. The oral examination revealed fissures on the dorsal tongue. In addition to the lack of expression of saliva from the submandibular ducts, as observed in the initial visit, no saliva was expressed from the parotid ducts. This was consistent with the sialometric values of 0 mL/5 min, as on the initial evaluation.

On rheumatologic clinical examination, asthenia and no pain were reported, there was absence of purpura or other systemic manifestations. The serology results were the same as in the first visit except for the RF. Even though the value of 8 IU/mL was within normal limits (<14 IU/mL), in this second occasion it was double the initial value.

On ocular examination, keratoconjunctivitis sicca was observed, as in the initial examination, and similar results were obtained for the Schirmer test, BUT, and corneal staining with 0.5% fluorescein, with the only difference that filaments could be observed. The results of the conjunctival staining with 1% lissamine green were unchanged in both eyes.

The follow-up minor labial salivary gland biopsy showed atypical diffuse infiltration by mononuclear cells of variable size with little cytoplasm and hromorphic and hyperchromatic nuclei, affecting the whole specimen with destruction of glandular architecture. Histomorphologic studies included the use of an immunohistochemical panel for the determination of the phenotype. The result of the immunohistochemistry was that the lymphocytes were CD20+, CD3−, CD5−, and CD23−. Evidence of kappa light chain restriction and positive staining for Bcl-2 were observed. Ki-67, a measure of cellular proliferation, was positive in <5% of neoplastic cells (Figure 1). The diagnosis of B-cell MALT lymphoma of low grade, was made. Because the patient did not present clinical signs or symptoms of alteration in the parotid glands, no image diagnostic tests were requested.

Due to the presence of a gastric acid–sensitive syndrome she was referred to gastroenterology for study. An endoscopy was performed, and the gastric biopsy showed chronic gastritis with presence of Hp. Antibacterial treatment with amoxicillin-clavulanic acid (1 g
twice daily) plus clarithromycin (500 mg twice daily) for 8 days and pantoprazol (40 mg daily) for 8 weeks was prescribed. The general evaluation of the patient, including computerized tomography and bone marrow biopsy, showed no additional evidence of disease. The endoscopy and gastric biopsy performed after treatment showed mild chronic gastritis and absence of Hp. A “watch and wait” policy was adopted. Local and systemic evaluation was performed every 6 months during the first year and then, once a year with no evidence of disease dissemination to date.

DISCUSSION

Development of NHL is the most serious complication of SS. Several histologic subtypes of NHL have been described in patients affected with this syndrome, most commonly MALT lymphoma. This type of lymphoma has characteristic clinicopathologic features and is rare in the oral cavity. It generally has a relatively indolent course and remains localized to the original site for prolonged periods of time. This sometimes leads to mistaking it for an inflammatory process. However, in extragastrointestinal locations, the clinical development and biologic behavior is heterogeneous and may compromise multiple sites of the mucosa. This occurrence is most probable in head and neck MALT lymphomas. Histologically, it is composed of morphologically heterogeneous small B cells, including marginal zone (centrocyte-like) cells, others resembling monocytoid cells, small lymphocytes, and scattered immunoblasts and centroblast-like cells. The neoplastic cells infiltrate around reactive B-cell follicles in a marginal

Fig. 1. Histomorphology and immunohistochemistry of MALT lymphoma in minor salivary gland follow-up biopsy. A, Hematoxylin-eosin (original magnification ×40); B, CD20 (positive) in most lymphocytes exhibiting a diffuse pattern; C, CD3 (negative) identifies scattered T cells of the lymphocytic infiltrate; D, kappa light chain restriction; E, Bcl-2 positive staining; F, Ki-67 positive expression in <5% of neoplastic cells (original magnification ×100).
zone distribution and spread into the interfollicular areas. The determination of the immunophenotype, as well as the presence of immunoglobulin or Bcl-2 gene rearrangements, aid in the diagnosis, especially when it is difficult to distinguish from reactive lymphocytic infiltration. It has been suggested that prolonged autoimmune inflammation, as in SS, or persistent antigenic stimulation caused by H. pylori and/or hepatitis C virus could play a role in the development of this type of lymphoma.

The labial salivary gland biopsy is one of the diagnostic tests for SS and aids in detection of lymphoma. The evaluation must be very thorough, especially when the infiltration of mononuclear cells is very marked and uniform. In this event the use of immunohistochemical techniques is necessary to determine the diagnosis.

In the present case, scheduled recall allowed early detection of the presence of a MALT lymphoma of the labial salivary glands. General evaluation of the patient performed after this diagnosis showed no evidence of disease dissemination. The biopsy performed during follow-up gastroscopy after anti-H. pylori treatment revealed mild chronic gastritis and absence of H. pylori. There is strong evidence of the association between gastric MALT lymphoma and infection by H. pylori, and remission can be obtained with appropriate antibacterial treatment. Such association has not been reported in salivary glands, although some authors have reported the remission of a case of parotid MALT lymphoma in a patient with SS after having received anti-H. pylori treatment because of chronic gastritis associated with H. pylori. The treatment suggested for disease localized in the head and neck is either surgery or radiotherapy. Systemic chemotherapy is not required to prevent recurrence, but periodic local and systemic evaluation for a prolonged period of time is necessary. However, some centers prefer the “watch and wait” approach in patients with localized disease, because it may be many years before evidence of disease progression occurs.

In the present case, the latter option was preferred because after general evaluation there was no additional evidence of disease.

CONCLUSION

Repeated examinations, including labial salivary gland biopsies as part of the SICCA protocol, enable standardized and thorough evaluation of disease progress in patients with SS. In the present case, this protocol allowed early detection of a MALT lymphoma in a patient that did not present with clinical signs related to a neoplastic lesion.

The authors thank Dr. Troy Daniels, from the University of California, San Francisco, for his collaboration in some of the immunohistochemical techniques and for providing some of the micrographs.

REFERENCES


